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# Morphometric and Genetic Variation in Three Populations of Indian Salmon (*Polydactylus plebeius*)

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# Abstract

Morphometric character analyses and RAPD was used to discriminate and ratify the status of three populations of Indian salmon, *Polydactylus plebeius* along the coromandel coast of India. Morphometric analyses showed a clear pattern of differentiation between the stocks and revealed the discreteness of two groups, southern stock (Pazhayar) and northern stock (Cuddalore). The univariate analysis of variance showed significant differences between means of the samples for most morphometric descriptors. A total of 1077 scorable bands were produced using all ten arbitrary primers in three populations. An un-weighted pair-group method with arithmetic mean (UPGMA) dendrogram was constructed based on genetic values to show the genetic relationship among the three populations. The genetic diversity (H) of *P. plebeius* in Cuddalore was more (0.0733 ± 0.0648) than Pazhayar (0.0609 ± 0.0416) and Vellar (0.0613 ± 0.0344) populations. All the three populations had significantly (p<0.001) higher interpopulation genetic distance value than the intrapopulation value. Further molecular studies, comprising more markers and populations are still required to precisely evaluate the genetic structure of threadfin fishes throughout the Indian coast.

Keywords: genetic variation, Indian salmon, morphometric, Polydactylus, population structure

# Introduction

Threadfin fish belongs to the family Polynemidae, which forms part of the order Perciformes. They typically inhabit marine coastal waters, estuaries and freshwater river mouths. This family has a wide distribution in the tropical parts of Atlantic, Indian and Pacific Oceans (Kagwade, 1970). The potential annual yield of polynemids in India is estimated around 9,000 tones (Srinath and Balan, 2003). Present catch of polynemids have gone down mainly because of the introduction of shrimp trawlers which has destroyed most of the young-ones of the larger varieties of polynemids grow to more than a meter (Prasad *et al.*, 2005).

Morphometric analysis has been applied to many stock differentiation and life-history problems in many fish species (Bronte *et al.*, 1999). If shape differences in different populations of the same species can be used to discriminate morphotypes, they may also be useful in examining the stock structure within a morphotype (Joseph and Jayasankar, 2001). Detection of differences within a morphotype may indicate geographically separated stocks, whose shapes may be predicted on local environmental conditions or have genetic bases (Joseph and Jayasankar, 2001). They discriminate two Nemipterus populations through morphometric and meristic characters in India. Turan *et al.* (2006) reported the genetic and morphological variation of *Pomatomus saltatrix* throughout the Black Seas, Marmara, Aegean and eastern Mediterranean Seas. Erguden *et al.* (2009) underwent morphometric and meristic analyses of chub mackerel *Scomber japonicus* to discriminate stocks throughout the Black, Marmara, Aegean, and northeastern Mediterranean Seas.

Genetic markers are generally oversensitive to a low level of gene flow, relatively low level of exchange between stocks, which are quite negligible from a management perspective, and may be sufficient to ensure genetic homogeneity (Carvalho and Hauser, 1994; Ward and Grewe, 1994). RAPDs have gained considerable attention particularly in population genetics (Lu and Rank, 1996), species and subspecies identification (Bardakci and Skibinski, 1994), phylogenetics, linkage group identification, chromosome and genome mapping, analysis of interspecific gene flow and hybrid speciation, and analysis of mixed genome samples (Hadrys *et al.*, 1992), breeding analysis and as a potential marker for single-locus genetic fingerprints (Brown and Epifanio, 2003).

In finfishes, size and morphometric variations among populations continue to play an important role in stock identification, despite the advent of biochemical and molecular genetics technique which help in identification of genetic differences between groups (Swain and Foote, 1999). Considering the importance of morphometric, genetic variation and as there are no attempts to study in *P. plebeius*, in spite of its significant contribution in Indian marine fish landing the present study was undertaking. The objective of this study was to evaluate patterns of morphological and genetic variation in the Indian salmon, *P.* 

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*plebeius* in three estuarine regions, by analyzing morphometric characters and banding pattern using ten arbitrary primers.

## Materials and methods

#### Sample collection

The striped threadfin, *P. plebeius* were collected from three stations such as, (S1) Cuddalore estuary (Lat. 11°42' N; Long. 74°46' E), (S2) Vellar estuary (Lat. 11°29' N; Long. 79°46' E) and (S3) Pazhayar estuary (Lat. 11°21' N; Long. 79°49' E) given in Fig. 1. Totally fifty individuals from each station were sampled for morphometric character measurements and tissue samples DNA isolation.

# Morphometrics

Forty nine morphometric characters were measured as per earlier report (Motomura *et al.*, 2001) and used for further analysis. The morphometric data were analysed using sheared PCA as per Bookstein *et al.* (1985). This technique, which quantifies shape differences independent of size, as previously been used to distinguish fish species (Browers and Stauffer, 1993). Morphometric measurements were log transferred to preserve allometrics, standardize variance and produce a scale invariant covariance matrix before analysis. To ensure comprehensive analyses of the data for more powerful discrimination between populations, sheared principal components of represented morphometric measurements were scattered against the first principal components in SPSS (V.14.0).

## DNA isolation and PCR

The DNA was isolated by standard Proteinase-K/Phenol-Chloroform-ethanol method (Sambrook *et al.*, 1989) and the concentration of isolated DNA was estimated using a UV spectrophotometer. The DNA was diluted in TE buffer to a final concentration of 100 ng/µL. Ten commercially available decamer random primers (An1-An10) from Chromous Biotech Pvt Ltd (Bangalore, India) were used for this study. The amplification reaction was carried out in a 25 µL reaction volume containing 10 mM Tris-HCl, 50 mM KCl (pH 8.5), 2.5 mM MgCl<sub>2</sub>, 0.001% gelatin, 100 µM of dNTP mix, 0.2 µM of each primer, 1 U of Taq DNA polymerase (Bangalore Genei, India) and 25 ng of template DNA. The PCR was performed in a thermocycle (TechGene, UK) for 40 cycles consisting of denaturation at 94°C for 30 seconds, annealing at 35°C for 30 seconds, and extension at 72°C for 60 seconds. The final extension was carried out at the same temperature for 5 minutes. The resulting products were electrophoretically analyzed through 1.5% agarose gels, stained with ethidium bromide, and visualized using a UV transilluminator. Subsequently the gel was photographed using a gel documentation system (Lark, India).

#### Data analysis

Genetic similarity/distance between the three stripped threadfin populations was estimated using Popgene Software (Version 1.31) (Yeh *et al.*, 1999). Nei and Li's (1979) genetic similarity (GS) among the three populations was computed and converted by Popgene into genetic distance (GD) according to Hillis and Mortiz's (1990) formula, GD = 1-GS. The GS reflects the proportion of the bands shared between individuals and values range from (1) when present to (0) when absent. Phylogenetic relationship was estimated based on genetic distance values generated from RAPD data among the three populations. A neighbor-joining dendrogram also generated based on Nei's procedure (Nei, 1978) using Popgene.

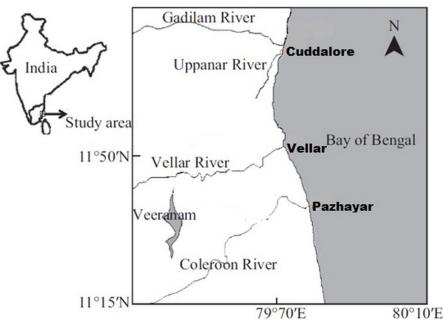


Fig. 1. Map showing the sample collection sites in Coromandel coast of India

Differences in intrapopulation and interpopulation genetic distance coefficients among the three populations were tested by one way analysis of variance (ANOVA). Intrapopulation and interpopulation genetic distance values were compared by paired *t*-test. The statistical analyses were performed by the software SYSTAT (version 7.0).

## Results

# Morphometrics

Morphometric values obtained from the three populations of *P. plebeius* are shown in the Tab. 1. Many of the characters in all the three populations are not significantly deviate. Vellar and Pazhayar populations are showing more morphological similar characters value when comparing to Cuddalore population. Except total length, fork length, maximum body height, overall caudal fin length, height and length of longest pectoral fin filament, the Cuddalore population also mingled with other two populations. The principal components analysis score plot also exhibited a similar trend of clustering of Cuddalore and Pazhayar population separately. But, the Vellar population individuals are shared with Pazhayar and Cuddalore populations (Fig. 2).

# Genetic diversity

By using the ten random primers (An1-An10) in three populations, totally 1077 scorable bands were observed. In Cuddalore population, total numbers of bands were 330 and in Pazhayar, Vellar it was 417 and 330 respectively. Nei's (1978) unbiased genetic distances and genetic similarity between three populations of *P. plebeius* are given in Tab. 2. The genetic distance between Pazhayar and Cuddalore was more (0.0034) than Vellar and Pazhayar (0.0033). The genetic identity between Cuddalore and Pazhayar was 0.9966, genetic identity between Pazhayar and Vellar was 0.9967 based on the RAPD data. The overall observed and expected polymorphic loci in three populations are given in Tab. 3. The genetic diversity (H) of *P. plebeius* in Cuddalore population was more  $(0.0733 \pm 0.0648)$  than Pazhayar  $(0.0609 \pm 0.0416)$  and Vellar  $(0.0613 \pm 0.0344)$ populations. The intrapopulation genetic distance values for the three populations were tested by one way ANOVA and found to be significantly slight different (p < 0.0001) (Tab. 4). But the interpopulation genetic distance values estimated for the three populations also tested by one way ANOVA and found to be significantly much different (*p*<0.001) (Tab. 4).

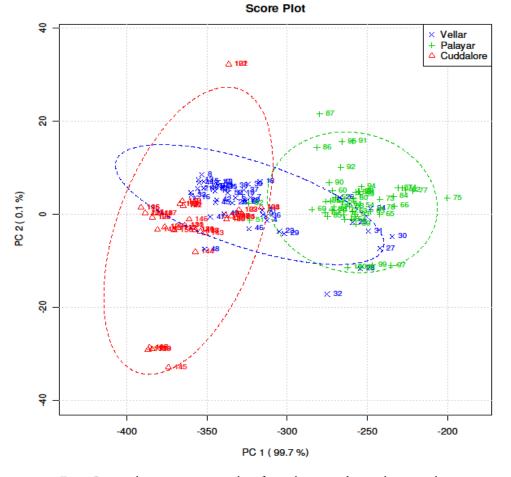


Fig. 2. Principal component score plot of morphometric data in three populations

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Tab. 1. Morphometric characters observed in three populat	ions of <i>P. plebeius</i>

No.	Morphometric characters (mm)	Vellar	Pazhayar	Cuddalore
1	TL	$130 \pm 15.20^{\circ}$	$156 \pm 28.1^{\circ}$	$187 \pm 11.63^{a}$
2	FL	$108.7 \pm 10.26^{a}$	125.8±30.32 <sup>b</sup>	152.3± 18.87ª
3	STL	98.14 ±11.59ª	114.4±21.64 <sup>b</sup>	133.5± 8.12ª
4	1 <sup>st</sup> P-DL	34.71±3.61 <sup>a,c</sup>	44.4±10.42ª	44.1±2.58 <sup>b,c</sup>
5	2 <sup>nd</sup> P-DL	61.51± 3.9 <sup>a</sup>	71.34±14.38 <sup>b</sup>	79.3±4.92ª
6	P-PTL	26.17±4.05 <sup>a,c</sup>	32.25±5.78ª	37.52±2.06 <sup>b,c</sup>
7	P-PVL	38.37±3.33 <sup>a,c</sup>	43.40±4.30ª	48.80±6.51 <sup>b,c</sup>
8	P-AL	63.4±5.80ª	72.96±12.94 <sup>b</sup>	85.66±4.94 <sup>a,b</sup>
9	HL	29.37±3.13 <sup>a,c</sup>	36.06±6.59ª	41.57±2.89 <sup>b,c</sup>
10	НН	17.97±2.8 <sup>a,c</sup>	28.06±7.20ª	28.14±1.79 <sup>b,c</sup>
11	HW	5.25±0.56 <sup>a,c</sup>	6.93±1.56ª	$7.66 \pm 0.85^{b,c}$
12	LMC	5.71±0.71ª,c	7.71±2.38ª	9.61±0.74 <sup>b,c</sup>
13	LMO	9.22±1.13 <sup>a,c</sup>	11.34±2.75ª	14±1.64 <sup>b,c</sup>
14	UJL	12.62±1.69 <sup>a,c</sup>	14.15±2.12 <sup>ª</sup>	14.95±0.49 <sup>b,c</sup>
15	MH	17.4±2.25ª	15.31±1.83 <sup>a,b</sup>	19.33±2.26 <sup>b</sup>
16	EH	12.59±1.17ª	14.56±2.15 <sup>a</sup>	18.61±1.56ª
17	OD	7.03±0.93ª	8.46±1.64ª	10.5±0.70ª
18	DEO	3.64±0.52 <sup>a,c</sup>	4.98±0.98ª	5.57±0.53 <sup>b,c</sup>
19	EA	21.64±2.83 <sup>a</sup>	24.78±5.42 <sup>b</sup>	32.61±2.37 <sup>a,b</sup>
20	MBH	30.02±2.36ª	35.21±6.72ª	43.04±3.52ª
21	MBW	5.45±0.70 <sup>a</sup>	6.53±1.39 <sup>b</sup>	5.80±0.60°
22	B 1st-DFL	13.85±1.71ª	16.37±3.56 <sup>b</sup>	19.19±1.99ª
23	B 2 <sup>nd</sup> -DFL	19.11±2.25 <sup>a,c</sup>	22.59±4.25°	25.14±1.82 <sup>b,c</sup>
24	B-AFL	19.82±1.82ª	21.40±3.67 <sup>b</sup>	24.04±1.80ª
25	B-PTFL	5.25±0.56ª	5.56±0.91 <sup>b</sup>	6.95±0.66 <sup>a,b</sup>
26	B-PVFL	4.71±0.51ª	4.56±0.71 <sup>b</sup>	5.19±0.40°
27	1st-DFH	19.4±2.58ª	23.31±4.50ª	28.61±2.29ª
28	2 <sup>nd</sup> -DFH	18.74±2.52ª	23.40±4.17 <sup>a</sup>	28.23±2.02ª
29	AFH	$17.4\pm 2.08^{\circ}$	20.68±3.88ª	25.33±1.90ª
30	PTFH	19.34±2.38ª	22.78±3.77 <sup>a</sup>	27.66±2.55ª
31	PVFH	14.82±1.38ª	16.12±2.87 <sup>b</sup>	20.85±1.52 <sup>a,b</sup>
32	L.L 1 <sup>st</sup> -DFR	19.4±2.58°	23.31±4.50°	28.61±2.29ª
33	L.L 2 <sup>nd</sup> -DFR	18.77±2.53ª	23.40±4.17 <sup>a</sup>	28.23±2.02ª
34	L.L-AFR	17.4±2.08ª	20.68±3.88ª	25.33±1.90ª
35	L.L-MIR L.L-PTFR	19.37±2.38ª	20.08±3.77 <sup>a</sup>	27.66±2.55 <sup>a</sup>
36	L.L-PVFR	17.5/±2.38 14.54±2.41ª	16.12±2.87 <sup>b</sup>	27.00±2.55 20.85±1.52 <sup>a,b</sup>
37 37	O 1 <sup>st</sup> -DFR-AFL	$41.82 \pm 4.05^{\circ}$	50.65±9.14 <sup>a</sup>	62.04±4.99ª
38	O 2 <sup>nd</sup> -DFR-AFL		35.84±6.57ª	43.28±3.22ª
38 39	O 2 -DFR-AFL O PVFR-AFL	$30.57\pm2.44^{a}$ $27.34\pm3.10^{a}$		45.28±5.22 38.28±4.45ª
	2 <sup>nd</sup> DF SPINE	7.37±1.37ª.c	31.75±4.57ª 9.65±2.29ª	
40				$10.66 \pm 1.06^{b,c}$
41	L.L AF SPINE	$6.68 \pm 1.49^{a}$	7.93±1.58 <sup>b</sup>	9.85±0.96 <sup>a,b</sup>
42	CPL	$11.05 \pm 1.02^{a}$	12.40±2.56 <sup>b</sup>	$12.57 \pm 1.36^{a}$
43	CPH	$13.31 \pm 1.40^{a,c}$	16.90±3.55ª	19.28±1.67 <sup>b,c</sup>
44	CPW	4.65±0.68ª	5.5±1.10 <sup>b</sup>	5.66±0.65ª
45	UCFL	36.28±3.34ª	40.75±7.96 <sup>b</sup>	50.23±3.61ª.b
46	LCFL	33.31±3.48ª	38.78±6.59ª	46.04±3.26ª
47	CFH	45.65±4.56 <sup>a,c</sup>	54.56±10.81ª	57.42±3.13 <sup>b,c</sup>
48	L.L PT F.FILAMENT	36.05±2.36 <sup>a</sup>	40.21±6.99 <sup>b</sup>	49.76±2.48 <sup>a,b</sup>
49	BMH	15.37 ±1.43ª	19.03 ±3.76 <sup>a</sup>	23.66 ±2.45 <sup>a</sup>

The common superscripts sharing the row is not significantly different (p < 0.001)

(TL: Total length; FL: Fork length; STL: Standard length; 1st. P-DL: 1st Pre-dorsal length; 2nd. P-DL: 2ndPre-dorsal length; P-PTL: Pre-pettoral length; P-PVL: Pre-pelvic length; P-AL: Pre-anal length; HL: Head length; HH: Head height; HW: Head width; LMC: Length of snout with mouth closed; LMO: Length of snout with mouth opened; UJL: Upper jaw length; MH: Mouth height; EH: Eye height; OD: Orbit diameter; DEO: Dermal eye opening; EA: Eye area; MBH: Maximum body height; MBW: Maximum body width; B. 1st-DFL: Base of 1st dorsal fin length; 2nd-DFL: Base of 2nd dorsal fin length; B-AFL: Base of anal fin length; B-PTFL: Base of pectoral fin length; B-PVFL: Base of pectoral fin height; CMC: Length of longest 1st dorsal fin height; 2nd-DFH: 2nd Dorsal fin height; AFH: Anal fin height; PTFH: Pectoral fin height; PVFH: Pelvic fin height; L. L. 1st-DFR: Length of longest 1st dorsal fin ray; L. L. 2nd-DFR: Length of longest 2nd dorsal fin length; of 1st dorsal fin ray; L. L-PTFR: Length of longest pectoral fin ray; L. L-PVFR: Length of longest pelvic fin ray; C. 1st DFL: AFR: Length of longest anal fin length; O. 2nd DFL-AFL: Origin of 2nd dorsal fin length to anal fin length; O. PVFL-AFL: Origin of pelvic fin length; 2nd dorsal fin spine length; CPU: Caudal peduncle length; CPH: Caudal peduncle height; CPW: Caudal peduncle width; UCFL: Upper caudal fin length; CPH: Caudal fin height; L. L. NTE. FILAMENT: Length of longest pectoral fin filament; BMH: Body mid line height)

Populations	Cuddalore	Pazhayar	Vellar
Cuddalore	****	0.9966	0.9966
Pazhayar	0.0034	****	0.9967
Vellar	0.0034	0.0033	****

Tab. 2. Nei's genetic identity (above diagonal) and genetic distance (below diagonal) of three populations of *P. plebeius* 

## Discussion

As per the present study, Vellar and Pazhayar populations are close enough when compared to Cuddalore population and exhibiting low phenotypic differentiation in PCA scatter plot analysis. The obtained *p*-values denote morphometric results are insignificant to support

Tab. 3. Overall observed number of alleles (Na), effective number of alleles (Ne), Nei's gene diversity (H), Shannon's information index (I), Number of polymorphic loci (Np) and Percentage of polymorphic loci (Pp) in three populations of *P. plebeius* 

Populations	Na	Ne	Н	Ι	Np	Pp (%)
Cuddalore	1.3347±0.4721	1.0447±0.0832	0.0733±0.0648	0.1173±0.0737	332	33.47
Pazhayar	$1.4183 \pm 0.4935$	1.0483±0.0777	$0.0609 \pm 0.0416$	0.1124±0.0839	415	41.83
Vellar	1.3327±0.4714	$1.0407 \pm 0.0804$	0.0613±0.0344	0.1107±0.0684	330	33.27
Overall	$0.0401 \pm 0.0007$	0.0379±0.0006	$0.0540 \pm 0.0317$	8.7672	1077	92.10

Tab. 4. Summary of results of one way ANOVA to test for differences in intrapopulation and interpopulation genetic distance value calculated based on RAPD markers among the three populations

Source of Variation	Sum of Squares	df	Mean squares	F	Р
Within Cuddalore population	1.0958	9	0.1217	12.2173	0.0001
Error	0.0985	58	0.0896		
Total	1.1943	67	0.2113		
Within Pazhayar population	1.4883	9	1.6537	10.2890	0.0001
Error	0.0894	62	0.8903		
Total	1.5777	71	2.5440		
Within Vellar population	1.5423	9	1.7137	10.2362	0.0001
Error	0.9225	55	0.6742		
Total	2.4648	64	2.3879		
Between three populations	2.9215	15	2.5632	34.4703	0.001
Error	1.1264	561	0.1633		
Total	4.0479	576	2.7265		

Based on Nei's genetic distance value, an UPGMA dendrogram was constructed and given in (Fig. 3). The cluster values indicated distinct relationship between the three populations of *P. plebeius*. In this dendrogram Pazhayar and Vellar populations are closely related than Cuddalore population.

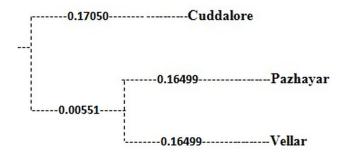


Fig. 3. Neighbour joining tree (1000 replications) generated from RAPD data of the three populations

the established differentiation between these three populations that often leads to taxonomic uncertainity. Some of the Pazhayar individuals clustered with Vellar population and some individuals clustered in a separate place in the plot. Only some individuals in Vellar population placed in between the Cuddalore and Pazhayar populations and remaining all the individuals dropped in a separate cluster in the plot.

The close distribution of these samples may be accounted for recent separation due to ecological alterations. Discrimination of the three populations as a single entity could be confirmed statistically by the insignificant difference observed from the morphometric data. Turan *et al.* (2006) studied the morphological variation of *Pomatomus saltatrix* based on morphometric and meristic analyses of samples collected throughout the Black Sea, Marmara, Aegean and eastern Mediterranean Seas and the results indicated existence of three morphologically differentiated groups. Erguden *et al.* (2009) underwent morphometric and meristic analyses of chub mackerel, *Scomber japonicus* throughout the Black, Marmara, Aege280

an, and northeastern Mediterranean Seas. They observed a clear pattern of morphometric and meristic differentiation between the stocks. The present study also showed a clear pattern of differentiation between the stocks and revealed two groups, the southern estuary stock (Pazhayar) and the northern estuary stock (Cuddalore).

The RAPD profile showed 92.10% polymorphic loci among the three estuarine populations in this study. As compared to previous studies, 68% of polymorphic loci were reported in three populations of catfish, Clarias batrachus in three water bodies of Bhopal in India (Mehrotra et al., 2010). Rahman et al. (2009) studied genetic variations of wild and hatchery populations of Catla catla revealed by RAPD markers and found overall 54.55% polymorphism. Shifat et al. (2003) have also reported RAPD Jaccards dissimilarity coefficient in 34 individuals of Tenualosa ilisha and observed 20.41% polymorphism. Rajasekar et al. (2012) investigated that the RAPD profile showed 93.12% polymorphic loci among the three populations of Lates calcarifer. The present study inferred from these RAPD data that showed relatively high level (92.10%) of genetic polymorphism and is possibly due to the small sampling size.

While considering the Nei's gene diversity (H), it was high  $(0.0733 \pm 0.0648)$  in Cuddalore population compared to the Vellar  $(0.0613 \pm 0.0344)$  and Pazhayar  $(0.0609 \pm 0.0416)$  populations, which are more or less similar. This result also coincides with previous study revealed by RAPD marker, where *Nemipterus nemurus*  $(0.0652 \pm 0.0520)$  showed similar gene diversity than *N. cyanomus*  $(0.0354 \pm 0.0208)$  and *P. caeruleus*  $(0.0283 \pm 0.0197)$  (Parveen *et al.*, 2011). Their results illustrate that genetic structure of damselfish population differ upon at the genus level not at the species level.

Nei's genetic identities between the three populations were high and it is evident that those three distinct populations were recently isolated or they are inhabited with weak geographical barriers. Similar genetic identity and genetic distance values were observed in the genetic diversity analysis of two Gangetic riverine populations of *Eutropiichthys vacha* using RAPD (Chandra *et al.*, 2010). They confirmed eventhough these two riverine populations vary spatially but originate from the same drainage system. In theory, the intrapopulation genetic distance values are expected to be lower than the interpopulation genetic distance. In this study also all the three populations had significantly (p<0.001) higher interpopulation genetic distance value than the intrapopulation value.

The clustering pattern obtained by UPGMA method both the Pazhayar and Vellar populations were fell in one clade and Cuddalore population displayed in another clade. This clustering pattern is directly proportional to the geographical distance. The same type clustering pattern regarding the geographic distance was obtained with the RAPD profiling analysis study on wild fish populations of *Catla catla, Eutropiichthys vacha, Clarias batrachus, Tenu*- *alosa ilisha* and *Lates calcarifer* (Shifat *et al.*, 2003; Rahman *et al.*, 2009; Chandra *et al.*, 2010; Mehrotra *et al.*, 2010; Rajasekar *et al.*, 2012).

# Conclusions

Statistical inference on morphometric data in this study revealed that it is insignificant to consider all three fish populations are widely distinct. RAPD gene diversity indices and genetic identity and distance values go accordingly same as mentioned with morphometric data. This was again corroborated with PCA and scatter plot analysis as well as dendrograms structured using UPGMA method. From the inferences claimed with previous studies in other fishes, it is feasible to consider all three populations as a single genetic structure. This presumption could be authenticated henceforth with other molecular markers. Further molecular studies, comprising more markers with other populations are still required to precisely evaluate the genetic structure of threadfin fishes along the Indian coast.

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