

Anti-Inflammatory Activity of *Salvadora persica* L. against Carrageenan Induced Paw Oedema in Rat Relevant to Inflammatory Cytokines

Abeer Y. IBRAHIM*, Souad E. EL-GENGAIHI, Hemaia M. MOTAWEA, Amany A. SLEEM

National Research Center, 12622 Cairo, Egypt; abeeryousry@yahoo.com (*corresponding author)

Abstract

In this study, the anti-inflammatory effect of aqueous alcoholic crude extract and ethyl acetate extract of miswak sticks (*Salvadora persica* L.) was investigated in carrageenan induced rat paw oedema in respect to immunological parameters. Adult male sapargue dawely rats were classified into four groups, group I received the vehicle (0.25% gum acacia solution), group II received crude aqueous alcoholic extract orally at 100 mg/kg, group III received ethyl acetate extract (100 mg/kg) orally and group IV received indomethacin (20 mg/kg) orally, and served as standard reference. The oedema was quantified by measuring the hind paw thickness immediately before subplantar injection, and at 1, 2, 3 and 4 h. Blood samples were withdrawn after the 4th hour of carrageenan induction, centrifuged and sera were used for analysis of pro-inflammatory cytokines. Administration of aqueous alcoholic extract and ethyl acetate extract (100 mg/ml) significantly reduced the oedema thickness in a time dependent manner, the inhibition percentage of inflammation was 17% for crude extract and 27% for ethyl acetate extract. Also the two extracts reduced secretion of inflammatory mediators, interleukin-1 β (IL-1 β), IL-6, tumor necrosis factor- α (TNF- α) and transforming growth factor- β 1 (TGF- β 1) in serum. The ethyl acetate extract shows potent activity to be nearly the same of indomethacin activity on all determined parameters at the last hour of following up. These results may be due to the presence of flavonoids in ethyl acetate extract. Three major flavonoids were isolated from ethyl acetate extract and identified as apigenin rhamnoglucoside, luteolin glucoside and rutin. The experimental study revealed that *Salvadora persica* extracts display remarkable anti-inflammatory activity.

Keywords: anti-inflammatory, cytokines, flavonoids, *Salvadora*, interleukin

Introduction

A large number of plant species contain various bio-active compounds exhibiting health beneficial properties; anti-oxidative, anti-inflammatory and mainly antimicrobial effects. Numerous natural products have been tested on various animal models for the development of new anti-inflammatory therapeutics (Sannigrahi, 2010).

Acute inflammation is rapid, short-lived and characterized by accumulations of fluid, plasma proteins and leukocytes. At the site of inflammations, the injured vascular endothelial cells and the emigrated leukocytes release a large number of soluble mediators which modulate and maintain the inflammation (Rosenberg and Gallin, 1999). Inflammation exerts multiple actions on the growth, differentiation and functions of lymphoid and non-lymphoid cells and regulates various aspects of the immune response and its production during injury or infection (Sarlis *et al.*, 1992). Chronic inflammation results in a progressive shift in type of cells present at site of inflammation and it is characterized by simultaneous destruction and healing of the injured tissue from incidence of inflammation. Carrageenan-induced mouse paw oedema has been used for assessment of the anti-inflammatory activity of many plant extracts and essential oils (Hajhashemi *et al.*, 2003; Khalil *et al.*; 2006; Orhan *et al.*, 2006). The toothbrush

tree, *Salvadora persica* L., locally called miswak, is a member of the *Salvadoraceae* family, has been used by many Islamic communities as toothbrushes and has been scientifically proven to be very useful in the prevention of tooth decay, even when used without any other tooth cleaning methods (Salehi and Momeni, 2006). Chewing sticks that are made from the roots, twigs, or stems of *S. persica* are commonly used in the Middle East as a means of maintaining oral hygiene. Studies indicate that *S. persica* extract is comparable to other oral disinfectants and anti-plaque agents, such as triclosan and chlorhexidine gluconate, if used at a very high concentration (Almas, 2002; Almas *et al.*, 2005). The present study aimed to investigate anti-inflammatory effect of *Salvadora persica* extract in oedma model focusing on four of anti-inflammatory mediators, namely; interleukin-1 β (IL-1 β), interleukin-6 (IL-6), tumor necrosis factor- α (TNF- α) and transforming growth factor- β 1 (TGF- β 1).

Materials and methods

Preparation of extracts

Salvadora persica plant was collected from Aswan Botanical Garden, Egypt, and was identified by the Botanist in Agriculture Dept. of the Botanical Garden. The fresh miswak sticks were cut into small pieces and allowed to

dry at room temperature ($30 \pm 2^\circ\text{C}$) for ten days then it was ground to powder in a ball mill. The powder (300 g) was exhaustively extracted with alcohol (70%) and then concentrated under reduced pressure using rotary evaporator to a small volume (100 ml). The crude ethanolic extract was fractionated using gradient solvents. Each extract was concentrated under reduced pressure to be free from solvents. The ethyl acetate extract and crude extract were used in this anti-inflammatory study. Chemicals, carrageenan and indomethacin, were purchased from Sigma-Aldrich GmbH, Sternheim, Germany.

Animals and experimental design of anti-inflammatory activity in Carrageenan induced paw oedema model

Carrageenan-induced rat paw oedema has been used for assessment of the anti-inflammatory activity of many plant extracts. The method used was carried out according to Winter *et al.* (1962).

Adult male sapargue dawely rats weighing between 150-180 g were maintained under normal laboratory conditions and kept in standard polypropylene cages at room temperature of $25-30^\circ\text{C}$, 60 to 65% relative humidity and provided with standard diet and water *ad libitum*. The experimental protocols were approved by institutional Animal Ethical Committee in National Research Center (registration no. 90123). Four groups each of eight rats were used and treated as follow:

Group I: received the vehicle (0.25% gum acacia solution) orally using orogastric tube and served as control group;

Group II: received crude aqueous alcoholic extract orally at 100 mg/kg;

Group III: received ethyl acetate extract (100 mg/kg) orally;

Group IV: received indomethacin (20 mg/kg) orally, and served as standard reference.

One hour following the previously mentioned treatments, paw swelling was induced by subcutaneous injection of 100 μl of 1% sterile lambda carrageenan suspension in saline into the right hind paw of all groups. The oedema was quantified by measuring the hind paw thickness immediately before subplantar injection, and at 1, 2, 3 and 4 h with a micrometer caliber. The increase in paw volume was calculated as the difference between the right paw volume (carrageenan) and the left paw volume (saline) measured at each time point. The percentage of inhibitory activity at 5th hour was calculated according to the following formula (Hajhashemi *et al.*, 2003):

$$\text{Percent Inhibition} = [(C - T) / C] \times 100$$

where, 'T' represents increase in oedema volume in animal's paw administered tested materials and 'C' represents increase in oedema volume in control.

Blood samples were withdrawn after the 4th hour of carrageenan induction, centrifuged at 3500 rpm. Sera aliquots were frozen at -80°C for analysis of pro-inflammatory cytokines.

Determination of Interleukins, tumor necrosis factor- α and transforming growth factor- β 1

IL-1 β and IL-6 levels were assayed in sera using enzyme-linked immunoadsorbent assay kits in accordance with the manufacturer's recommendations (Quantikine, USA), tumoral necrosis factor- α (Hycult biotech, Netherlands) while transforming growth factor- β 1 (Abnova, Taiwan). The assessment was done by ELISA reader (NJ 2000; Nihom Inter Med. Co), the sensitivity of assay was 20 pg/ml.

Statistical analysis

Data are expressed as mean \pm standard deviation or Standard error, and they were analyzed by the one-way analysis of variance (ANOVA), followed by post-hoc Tukey's test for multiple comparisons. The differences were considered significant if the probability was associated with $p < 0.01$.

*Isolation and purification of major flavonoids from *S. persica* ethyl acetate extract*

As crude alcoholic and ethyl acetate extracts showed anti-inflammatory activity, both extracts were subjected to phytochemical examination using magnesium turning test (Mabry *et al.*, 1970) which proved the occurrence of flavonoids.

The purification of major flavonoids was carried out according to Mabry *et al.* (1970). Ethyl acetate fraction was used for isolation of *Salvadora persica* main flavonoids using preparative TLC (GF254) technique. Ethyl acetate: methanol (9:1), ethyl acetate: methanol: water (6:2:1) and acetic acid 15% were used for fractionation as solvent systems. The isolated compounds were further purified on sephadex (LH-20) column using butanol saturated with water as solvent system.

The purified compounds were analyzed using UV spectroscopy (Shimadzu UV-240IPC) with different reagents; sodium methoxide, sodium acetate, sodium acetate with boric acid, aluminum chloride and aluminum chloride with hydrochloric acid. The purified flavonoids were subjected to $^1\text{H-NMR}$ (Joel ECA, 500MHz), $^{13}\text{C-NMR}$ (Joel ECA, 125MHz) and mass-spectroscopy (Finningen, model 3200 at 70eV). The sugar moieties of the isolated flavonoid glycosides were investigated according to Gálvez *et al.* (2003).

Results

*Effect of *S. persica* extracts and indomethacin on oedma in Carrageenan model*

Carrageenan-induced paw edema as an *in vivo* model of inflammation is the most frequently used method to evaluate the anti-edematous effect of natural products. The results obtained are summarized in Tab. 1.

Carrageenan injection induced skin inflammation in rat hind paw which increased paw diameter to be 4.32

mm, however, administration of *Salvadora* extracts significantly reduced the inflammation in hind paw time dependently (Fig. 1). At the first hour, administration of *Salvadora* extracts didn't show any significant reduction in oedema as compared to vehicle control group or reference drug. Through the following up period, the effect of ethyl acetate was increased in an ascending order to be similar to indomethacin at the last hour while crude extract remained the lowest effective one. The reduction in paw diameter was 27.08 and 27.31% for ethyl acetate extract and reference drug, respectively after four hours of carrageenan administration. It could be concluded that the ethyl acetate extract is more potent than the crude extract in treating oedema.

Effects of *S. persica* extracts and indomethacin on serum inflammatory cytokines concentration

In assay data, the IL-1 β , IL-6, TNF- α and TGF- β 1 secretion in serum were highly elevated by carrageenan induction but administration of *S. persica* extracts signifi-

cantly reduced serum secretion of inflammatory mediators as compared to vehicle group (Tab. 2).

IL-1 β was elevated in response to carrageenan induction (48.2 ng/l) inversely with extract pretreatment which caused a significant reduction in IL-1 β secretion from 48.02 ng/l in vehicle control group sera to 33.53 and 29.77 ng/l for animals of crude and ethyl acetate extracts groups, respectively. The same effect was observed in IL-6 secretion as the two extracts produced depletion percentages of 10.5 and 19.5% for crude and ethyl acetate extracts, respectively and 19.8% for indomethacin administration. Data presented in Tab. 2 clearly indicate that ethyl acetate extract has similar effect to indomethacin on IL-1 β and IL-6 with insignificant difference between them.

TNF- α secretion was magnified by carrageenan injection (371 pg/ml). TNF- α secretion was reduced, comparable to carrageenan group, as a respond to extract administration. Administration of ethyl acetate extract significantly reduced inflammatory mediator (TNF- α) secretion by 28.9% to be close to inhibition level of indomethacin administration (32.2%). Secretion of TGF- β 1

Tab. 1. Acute inflammatory activity of *S. persica* extracts and indomethacin (reference drug) on Carrageenan induced paw oedema in rats

Time in hours	Paw diameter (mm)				
	Zero time	1 h	2 h	3 h	4 h
Groups	Oedema inhibition (%)				
Control vehicle (Carrageenan+1 ml saline)	2.66±0.09	3.66±0.07*	3.75±0.13*	3.9±0.13*	4.32±0.10*
Crude extract (100 mg/kg)	3.13±0.11	3.85±0.18*	3.77±0.14*	3.61±0.13* (7.4%)	3.56±0.14* (17.59%)
Ethyl acetate (100 mg/kg)	2.88±0.07	3.64±0.03* (0.55%)	3.37±0.01* (10.13%)	3.25±0.1* (16.66%)	3.15±0.06* (27.08%)
Indomethacin (20 mg/kg)	3.00±0.09	3.37±0.04* (10.13%)	3.28±0.06* (12.53%)	3.16±0.01* (18.97%)	3.14±0.01* (27.31%)

Data are presented as mean \pm standard error followed by oedema inhibition percentage, * P <0.01

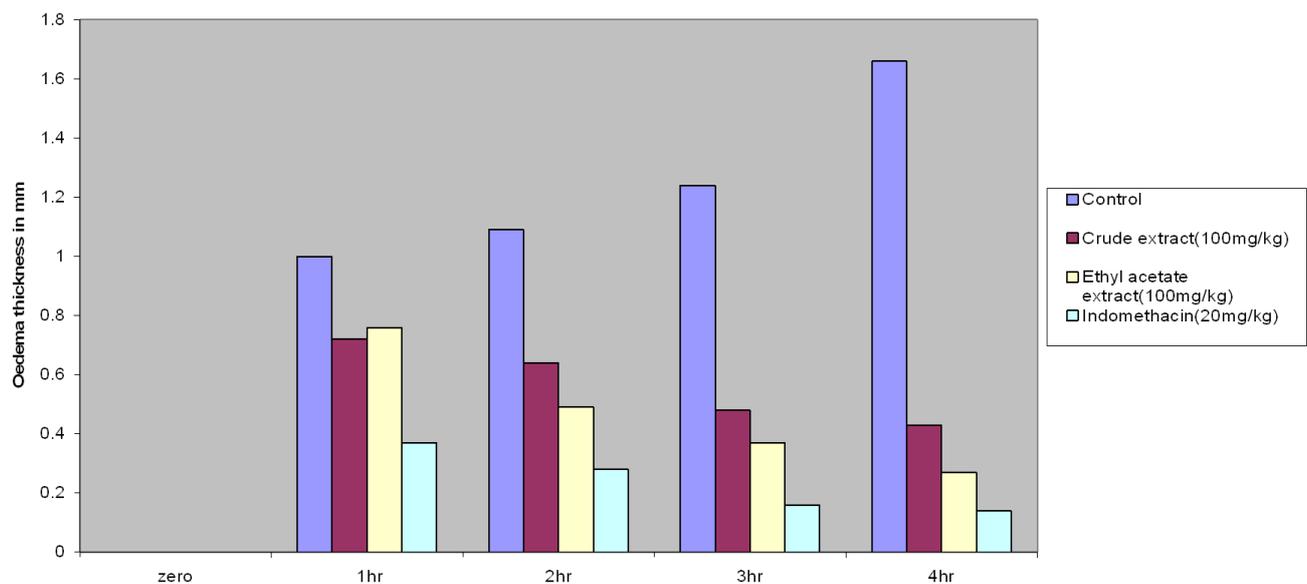


Fig. 1. Effect of *Salvadora persica* extracts on rat skin oedema thickness in carrageenan model. Values are mean of n, n=8

Tab. 2. Effect of *S. persica* on serum inflammatory cytokines secretion in carrageenan induced paw oedema in rat

Parameter Group	Interleukin-1 β (ng/l)	Interleukin-6 (pg/ml)	Tumor necrosis factor- α (pg/ml)	Transforming growth factor- β 1 (pg/ml)
	Inhibition (%)			
Control (v)	17.83 \pm 0.26 ^a	211.52 \pm 0.99 ^a	224.92 \pm 1.11 ^a	30.82 \pm 0.66 ^a
Control (Carrageenan)	48.02 \pm .58	405.70 \pm 4.37	371 \pm 0.77	100.69 \pm 4.49
Crude extract (100 mg/kg)	33.53 \pm 0.72 ^a (29.9%)	363.13 \pm 9.43 ^a (10.5%)	290.71 \pm 2.48 ^a (21.6%)	74.15 \pm 1.35 ^a (26.4%)
Ethyl acetate extract (100 mg/kg)	29.77 \pm 0.51 ^a (38%)	326.62 \pm 1.11 ^{ab} (19.5%)	263.61 \pm 1.05 ^a (28.9%)	61.33 \pm 0.85 ^{ac} (39.1%)
Indomethacin (20 mg/kg)	26.80 \pm 0.34 ^a (44.2%)	325.15 \pm 0.54 ^{ab} (19.6%)	251.36 \pm 4.92 ^a (32.2%)	59.52 \pm 6.51 ^{ac} (40.9%)

Data are presented as the mean \pm S.D. a $P < 0.01$, compared to vehicle control group; Groups have the same letter means do not differ significantly

reached a high concentration in serum by carrageenan induction however, administration of the two extracts decreased TGF- β 1 concentration in serum from 100.69 pg/ml in vehicle control group to be 61.33 pg/ml and 74.15 pg/ml in case of ethyl acetate and crude extracts, respectively. On the other hand, indomethacin reached nearly the same value (59.51 pg/ml).

Identification of *S. persica* isolated flavonoids

Purification of the ethyl acetate extract gave three main flavonoids, each of them showed a single spot with different solvent systems. Using acetic acid (15%) as solvent system, the R_f 's values of the purified three compounds were 0.51, 0.57 and 0.19 corresponding to apigenin rhamnoglucoside, luteolin glucoside and rutin, respectively.

Compound 1 (Apigenin rhamno-glucoside)

The UV spectral data of compound 1 indicated that this compound is flavone glycoside as it gave two absorption bands at 268 nm and 333 nm and it was glycosylated at C-7 position on A-ring. A bathochromic shift of band I from 333 nm to 384 nm with addition of sodium methoxide and shift with aluminum chloride to 380 nm. Also a shoulder at 340 nm indicates the presence of free hydroxyl groups at C-5 and C-4' positions. The bathochromic shift from 333 nm to 386 nm and 353 nm by addition of sodium acetate indicates free hydroxyl group at C-4' position with non free hydroxyl group at C-7 position. Therefore, the expected flavonoid structure is flavone with two free hydroxyl groups at C-5 and C-4' position and was glycosylated at C-7 position. Partial hydrolysis of this compound showed that it has two sugars; rhamnose and glucose. It is obvious from the above data that this compound is apigenin glycoside glycosylated with rhamnose and glucose, this expectation is coincided with data reported by Mabry *et al.* (1970) and Dordevic *et al.* (2000).

This expectation was also supported by ¹H-NMR, IR and Ms spectral data which showed four characteristic signals of flavone aromatic ring at 6.2 ppm, 7.5 ppm, 7.685 ppm and 8.52 ppm for H-6, H-5', H-3' and H-6',

respectively. The carbonyl group of C-ring gave IR band at 1700 cm⁻¹ and characteristic signal of rhamnose methyl group at 1.085 ppm to 1.233 ppm which gave a sharp band at 1416.5 cm⁻¹ in IR chromatogram. The recorded protons signals of rhamnose were as follow; signal at 3.282 for H-5'', 3.556 ppm for H-2'', 3.826 ppm for H-6'' and 4.6 ppm for H-1''. Signals at 3.11 ppm are for H-2 of glucose while band at 3424.2 cm⁻¹ on IR is for glucose hydroxyl groups. The UV spectral data and ¹H-NMR spectra are in accordance with Mabry *et al.* (1970) and El Alfy *et al.* (2010). It could be suggested that this compound is apigenin rhamno-glucoside.

Compound 2 (Luteolin glucoside)

The UV spectra of this compound suggested the presence of luteolin glucoside with a single sugar position. Compound 2 has two characteristic bands at 349 and 253 nm. The bathochromic shift with AlCl₃ produced two peaks at 426 and 328 nm indicating the presence of free OH in position 5, also the band shift with NaOAc+H₃BO₃ at 370 nm indicating the presence of free OH groups at 3 and 4 positions (Mabry *et al.*, 1970; Meririnhos *et al.*, 2005). Comparison of NMR spectra of compound 2 with those published by Wang *et al.* (1998) and Lu and Foo (2000), compound 2 could be identified as luteolin-7-O- β -D-glucoside. The doublet at δ 5.4 (J 1.9Hz) for anomeric proton in ¹H-NMR data indicating a β -configuration for glucose moiety. Compound 2 showed a singlet at 6.7 ppm in ¹H-NMR spectra consistent with the H-3 of flavone and this was supported by the observation of carbon signals at 103 ppm associated with the C-3 in the ¹³C-NMR spectra. ¹³C-NMR spectrum displayed C-5, C-7 and C-9 in the upfield regions at δ 146.7, 152.1 and 157, respectively, which agreed with 5, 7-hydroxylated A-ring of flavone (Horie *et al.*, 1998). The linkage between the sugar H-1'' (δ 5) and the C-7 position of the flavone is occurred in the upfield region at δ 152.8, (Tab. 3). Samples were analyzed in DMSO, 500 MHz exhibited molecular ion at m/z 446.5 whose fragmentation provided a characteristic m/z at 285, a typical mass for luteolin aglycone.

Compound 3 (Rutin)

The UV spectrum of compound 3 showed two major absorption bands at 359 and 257 nm, which indicated the presence of flavonol structure. The bathochromic shift with sodium methoxide supported the presence of 4'-hydroxyl group and with sodium acetate indicated the 7-OH function, while AlCl_3 and AlCl_3/HCl spectrum showed 5-OH and ortho di-OH in B-ring. The above data mean that 3-OH is absent or substituted. These results were supported by $^1\text{H-NMR}$ and $^{13}\text{C-NMR}$. The chemical shifts of protons and carbons essentially identical with those reported for rutin by Fathiazad *et al.* (2006) and Brietmaier and Voelter (1989). Also chemical shift at 6.21 ppm ($J=2$) was for proton of C-6 while shift at 6.4 ppm ($J=2$) for C-8 proton. The chemical shifts at 7.6, 6.81, 7.56 ppm for protons of C-2, C-5 and C6, respectively. The OH group on C-3, C-4, C-5 and C-7 showed chemical shifts at 9.1, 9.74, 12.6 and 10.9 ppm. The characteristic signal of β -D-glucose was at 5.21 ppm while the characteristic signal of H-1 rhamnose and methyl group were at 5.12 and 1.24 ppm respectively. $^{13}\text{C-NMR}$ readings (Tab. 3) were identical with reported data by Fathiazad *et al.* (2006) also Brietmaier and Voelter (1989).

Discussion

The present study was conducted to investigate the possible anti-inflammatory effects of *S. persica* extracts in carrageenan model relevant to their ability to block leukotriene secretion in animal serum. Effort was also directed towards the isolation and identification of the chemical constituents of the biologically active extract.

Paw swelling is one of the major factors in assessing the degree of inflammation and efficacy of the tested drugs (Begum and Sadique, 1988; Mizushima *et al.*, 1972). The crude and the ethyl acetate extracts of *S. persica* were investigated for their acute anti-inflammatory activity using oedema model. The two extracts significantly inhibited oedema in a time dependent manner and the maximum inhibition percentages were recorded with ethyl acetate extract. In addition, ethyl acetate extract reached the same inhibition level of the reference drug indomethacin at end of the follow up period.

Plasma levels of IL-6 appear to reflect the changes in inflammation (Gadiant and Otten, 1995) and it is an important immune mediator co-ordinating the activity of different immune cells with an important role in acute phase response (Harbuz *et al.*, 1993). Administration of *S. persica* extracts reduced the elevation of IL-6 by carrageenan induction. However, the ethyl acetate extract showed similar effect to the reference drug indomethacin, meaning that it is a potent agent in reducing inflammation.

Tumor necrosis factor α (TNF- α) is a pleotropic inflammatory cytokine produced by the immune system that suppresses tumor cell proliferation. Subsequent studies established that TNF- α is a key mediator of inflamma-

tion (Vasanthi *et al.*, 2007; Warren *et al.*, 1988,) so, it is an important parameter that should be included in determining the anti-inflammatory activity of plant extracts. Also, interleukin-1, an important cytokine produced mainly by blood monocytes, mediates the panoply of host reactions collectively known as acute phase response also known as Endogenous Pyrogen (EP), mononuclear cell factor and lymphocyte activating factor. TNF- α and IL-1 β are potent proinflammatory cytokines capable of inducing multiple signaling cascades that can serve in host defense and paradoxically contribute to inflammatory tissue injury (McCulloch *et al.*, 2006). Both IL-1 α and IL-1 β can trigger fever by enhancing prostaglandin E2 (PGE2) synthesis by the vascular endothelium of the hypothalamus and can stimulate T cell proliferation (Limeili *et al.*, 2008). Results of the present study revealed that the effects of *S. persica* extracts on IL-1 β and TNF- α were more pronounced than their effects on IL-6. Both of the two extracts significantly reduced the secretion of IL-1 β and TNF- α . The represented effect of ethyl acetate extract was similar to the standard drug indomethacin.

TGF- β distributed in tissues and synthesized by many different cells (Kropf *et al.*, 1997). It is involved in wound repair processes and in starting inflammatory reaction and then in the resolution through chemotactic attraction of inflammatory cells and fibroblast (Lawrance, 1996). Administration of extracts before carrageenan induction appeared to be preferable; they decreased TGF- β concentration in serum. The ethyl acetate extract was superior to crude one and gave the same results of indomethacin with insignificant difference between them.

These findings show that the ethyl acetate extract was able to inhibit acute inflammatory mediator release from mast cells and leukocytes, also to reduce inflammation by inhibition of TGF-1 which might participate in the early inflammatory responses (Giri *et al.*, 1993; Pittet *et al.*, 2001). The obtained results suggest that *Salvadora persica* ethyl acetate extract may decrease the vascular permeability, promote the resolution of oedema and exhibit inhibitory effect on proinflammatory cytokines secretion. Our results are in coincided with those of Saahil *et al.* (2007), Guardia *et al.* (2001) and Rajesh *et al.* (2011) who reported that *Salvadora persica* roots possess anti-oxidant activity and anti-inflammatory activity. Also *S. persica*, toothbrush stick, is used for oral hygiene, and to treat gum inflammation (Sher *et al.*, 2010).

Biochemical investigation carried out through this investigation revealed the presence of apigenin rhamnoglucoside, luteolin glucoside and rutin as the major flavonoids in the ethyl acetate extract of *S. persica*. Many plant derived compounds have been tested for their anti-inflammatory activity as ability to block leukotriene synthesis in cells from rat, mouse or human sources. Some plant derived chemical constituents like flavanoids, coumarins, quinones, pentacyclic triterpenes, sesquiterpenes, alkaloids and polyacetylates have been reported to be 5-Lipoxygenase

inhibitors (Sridhar *et al.*, 2004; Werz, 2007). Extracts of *Salvadora persica* possess various antiplaque, antiperiopathic, anti-inflammatory and antimycotic effects (Almas, 1993; Darout *et al.*, 2000). Rinsing with slurry of Miswak toothpaste reduces gingival inflammation and bleeding on probing. Also, *S. racemosa* exhibits a wide range of biological activities like antimicrobial, anti-oxidant, anti-lipidogen, anti-ulcer, anti-inflammatory, hepatoprotective, anti-diabetic (Al-Khateeb *et al.*, 1991). Flavonoidal glycosides which were isolated from this plant may be responsible for its pharmacological activities (Arora and Gupta, 2011). Several reports have suggested that diseases associated with oxidative stress as inflammatory diseases may be beneficially influenced by flavonoids (Rice-Evans *et al.*, 2000) because flavonoids influence a diverse range of intracellular signaling events depending on different features in the basic structure of flavonoids (Wang and Joseph, 1999).

Several mechanisms of action have been proposed for the bioflavonoid actions in reducing inflammation. Apigenin inhibits the collagenase activity involved in rheumatoid arthritis (RA) and suppresses lipopolysaccharide (LPS)-induced nitric oxide (NO) production in RA. Pretreatment with apigenin also attenuates LPS-induced cyclooxygenase-2 (COX-2) expression which is elevated in the synovial lining layer, the subsynovial synoviocytes, the vascular endothelial cells and mononuclear inflammatory cells in patients with RA or osteoarthritis (OA) (Lee *et al.*, 2007). In addition, apigenin profoundly reduces the tumor necrosis factor- α (TNF- α)-induced adhesion of monocytes to human umbilical vein endothelial cell (HUVEC) monolayer which is elevated in early RA (Lee *et al.*, 2007). It has been suggested that intradermal application of apigenin-7-glucoside inhibited skin inflammation caused by xanthine-oxidase and cumene hydroperoxide (Fuchs and Milbradt, 1993). Considering importance of luteolin, it may be a potent selective inhibitor of COX-2 and the inhibition is attributable to its down-regulation of the mRNA expression of COX-2 in inflammatory responses (Ziyan *et al.*, 2007). It inhibits arachidonic acid- and 12-O-tetradecanoylphorbol-13-acetate-induced ear edema (Ueda *et al.*, 2002). Luteolin and its derived glycosides have inhibitory activity against enzymes for the synthesis of thromboxane and leukotriene involved in inflammation (Odontuya *et al.*, 2005).

Conclusions

In conclusion, *Salvadora persica* extracts have anti-inflammatory properties. The potential efficacy of *S. persica* extracts to treat inflammation is based in a part on the hypothesis that it will suppress the proinflammatory cytokines resulting in less oedema.

References

- Al-Khateeb TL, OMullane DM, Whelton H, Sulaiman MI (1991). Periodontal treatment needs among Saudi Arabian adults and their relationship to the use of the Miswak. *Comm Dent Health* 8:323-28.
- Almas K (1993). Miswak and Its Role in Oral Health. *Post Grad Dent* 3:214-219.
- Almas K (2002). The effect of *Salvadora persica* extract (miswak) and chlorhexidine gluconate on human dentin: A SEM study. *J Conten Dent Pract* 3:27-35.
- Almas K, Skaug N, Ahmad I (2005). An *in vitro* antimicrobial comparison of miswak extract with commercially available non-alcohol mouth rinses. *Int J Dent Hyg* 3:18-24.
- Begum VH, Sadique J (1988). Long term effect of herbal drug *Withania somnifera* on adjuvant induced arthritis in rats. *Indian J Exp Biol* 26:877-882.
- Brietmaier E, Voelter W (1989). Carbon 13 NMR spectroscopy, 1st Ed. VCH Verlagsgesellschaft mbH, NY, 450-457 p.
- Darout IA, Christy AA, Skaug N, Egeberg PK (2000). Identification and quantification of some potentially antimicrobial anionic components in Miswak extract. *Ind J Pharmacol* 32:11-14.
- Dordevic S, Cakić M, Amr S (2000). The extraction of apigenin and luteolin from the sage *Salvia officinalis* from Jordan. *The Sci J Facta Univ, Series: Work Living Environ Protect* 1(5):87-93.
- El Alfy TS, El Sawi SE, Sleem A and Moawad DM (2010). Investigation of flavonoidal content and biological activities of *Chorisia insignis* Hbk. leaves. *Austr J Basic Appl Sci* 4(6):1334-1348.
- Fathiazad F, Delazar A, Amiri R, Sarker SD (2006). Extraction of flavonoids and quantification of rutin from waste tobacco leaves. *Iran J Pharmac Res* 3:222-227.
- Gadient RA, Otten U (1995). Identification of interleukin-6(IL-6) expressing neurons in the cerebellum and hippocampus of normal adult rats. *Neurosci Lett* 194:17-20.
- Gálvez M, Mart'ín-Cordero C, López-Lázaro M (2003). Cytotoxic effect of *Plantago* spp. on cancer cell lines. *J Ethnopharmacol* 88:125-130.
- Giri SN, Hyde DM, Hollinger MA (1993). Effect of antibody to transforming growth factor beta on bleomycin induced accumulation of lung collagen in mice. *Thorax* 48:959-966.
- Hajhashemi V, Ghannadi A, Sharif B (2003). Anti-inflammatory and analgesic properties of the leaf extracts and essential oil of *Lavandula angustifolia* Mill. *J Ethnopharmacol* 89:67-71.
- Harbuz MS, Rees RG, Lightman SL (1993). Hypothalamic-pituitary responses to acute stress and changes in circulating glucocorticoids during chronic adjuvant-induced arthritis in the rat. *J Physiol* 264:R179-185.

- Horie T, Ohtsura Y, Shihata K, Yamashita K, Tsukayama M, Kawamura Y (1998). ¹³CNMR spectral assignment of the A-ring of polyoxygenated flavones. *Phytochem* 47:865-874.
- Khalil NM, Sperotto JS, Manfron MP (2006). Antiinflammatory activity and acute toxicity of *Dodonaea viscosa*. *Fitoterapia* 77:478-480.
- Kropf J, Schurek JO, Wollner A, Gressner AM (1997). Immunological measurement of transforming growth factor-beta 1(TGF-1) in blood: assay development and comparison. *Clin Chem* 43(10):1965-1974.
- Lawrance DA (1996). Transforming growth factor-beta: A general review. *Eur Cytokine Netw* 7(3):363-374.
- Limeili ZF, Jianke R, Ruilin S, Zhihui L, Zhejin S, Long W, Xia S, Jun Y, Zhugang W, Jian F (2008). Functional imaging of interleukin 1 beta expression in inflammatory process using bioluminescence imaging in transgenic mice. *BMC Immunol* 9:49-54.
- Lu Y, Foo LY (2000). Flavonoid and phenolic glycosides from *Salvia officinalis*. *Phytochem* 55:263-267.
- Mabry TJ, Markham KR, Thomas MB (1970). The systematic identification of flavonoids, 2nd Ed. Springer Verlag, New York-Heidelberg-Berlin.
- McCulloch CA, Downey GP, El-Gabalawy H (2006). Signalling platforms that modulate inflammatory response: New targets for drug development. *Nature Rev Drug Discov* 5:864-876.
- Meririnhos J, Branca MS, Valenta PC, Seabra RM, Pepriera JA, Alberto D, Paula BA, Federico F (2005). Analysis and quantification of flavonoidic compounds from Portuguese olive (*Olea europaea* L.). *Nat Prod Res* 19(2):189-195.
- Mizushima Y, Tsukada W, Akimoto T (1972). A modification of rat adjuvant arthritis for testing antirheumatic drugs. *J Pharm Pharmacol* 24:781-785.
- Orhan DD, Küpeli E, Yesilada E, Ergun F (2006). Anti-inflammatory and antinociceptive activity of flavonoids isolated from *Viscum album* ssp. *album*. *Z Naturforsch C* 61:26-30.
- Pittet JF, Griffiths MJD, Geiser T, Kaminski N, Dalton SL, Huang XZ (2001). TGF-1 is a critical mediator of acute lung injury. *J Clin Invest* 107:1537-1544.
- Rosenberg HF, Gallin JI (1999). Inflammation In: *Fundamental Immunology*, p. 1051-1066. In: Paul WE (Ed.). Lippincott-Raven. 4th Philadelphia, New York.
- Salehi P, Momeni Danaie SH (2006). Comparison of the antibacterial effects of persica mouthwash with chlorhexidine on streptococcus mutans in orthodontic patients. *DARU* 14:178-182.
- Sannigrahi S, Parida S, Jagannath Patro V, Mishra US, Pathak A (2010). Antioxidant and anti-inflammatory potential of *Pterospermum acerifolium*. *Int J Pharm Sci Rev Res* 2(1):1-5.
- Sarlis NJ, Chowderly HS, Stephanou A, Lightman SL (1992). Chronic activation of the hypothalamo-pituitary-adrenal axis and loss of the circadian rhythm during adjuvant-induced arthritis in the rat. *Endocrinol* 130:1775-1779.
- Sridhar C, Subbaraju GV, Venkateswarulu Y, Raju TV (2004). New acylated iridoid glucosides from *Vitex altissima*. *J Nat Prod* 67:2012-2016.
- Vasanthi P, Nalini G, Rajasekhar G (2007). Role of tumor necrosis factor-alpha in rheumatoid arthritis: A review. *APLAR J Rheumatol* 10:270-274.
- Wang M, Li J, Rangarajan M, Shao Y, La Voie EJ, Huang TC, Ho TC (1998). Antioxidative phenolic compounds from sage (*Salvia officinalis*). *J Agricult Food Chem* 46:4869-4873.
- Wang H, Joseph JA (1999). Quantifying cellular oxidative stress by dichlorofluorescein assay using microplate reader. *Free Radic Biol Med* 27:612-616.
- Warren JS, Ward PA, Johnson KJ (1988). Tumor necrosis factor: A plurifunctional mediator of acute inflammation. *Modern Pathol* 1:242-247.
- Werz O (2007). Inhibition of 5-Lipoxygenase product synthesis by natural compounds of plant origin. *Planta Medica* 73:1331-1357.
- Winter C, Risley EA, Nuss GW (1962). Carrageenin induced oedema in hind paw of the rats for anti-inflammatory drugs. *Proc Soc Exp Biol Med* 3:544-547.